A Sysmex Group Company

CytoCell

Instructions For Use (IFU)

REF: CE-LPH 064-S / CE-LPH 064

FAST PML/RARα (RARA) Translocation, Dual Fusion Probe

CE IVD
2797

UK CA

PROFESSIONAL USE ONLY

ogt.com/IFU

Further information and other languages available at ogt.com/IFU

Intended Purpose

The CytoCell® *FAST* PML/RARα (RARA) Translocation, Dual Fusion Probe is a qualitative, non-automated, fluorescence *in situ* hybridisation (FISH) test used to detect chromosomal rearrangements between the 15q24 region on chromosome 15 and the 17q21.1-q21.2 region on chromosome 17 in Carnoy's solution (3:1 methanol/acetic acid) fixed haematologically-derived cell suspensions from patients with confirmed or suspected acute myeloid leukaemia (AML).

Indication for Use

This device is designed as an adjunct to other clinical and histopathological tests in recognised diagnostic and clinical care pathways, where knowledge of *PML::RARA* translocation status would be important for clinical management.

Limitations

This device is designed to detect rearrangements with breakpoints in the region covered by the red and green clones in this probe set, which includes the *PML* and *RARA* regions. Breakpoints outside this region, or variant rearrangements wholly contained within this region, may not be detected with this device.
This device is not intended for: use as a stand-alone diagnostic, use as a companion diagnostic, prenatal testing, population-based screening, near-patient testing, or self-testing.
This device has not been validated for sample types, disease types, or purposes outside of those stated in the intended purpose.
It is intended as an adjunct to other diagnostic laboratory tests and therapeutic action should not be initiated on the basis of the FISH result alone.
Reporting and interpretation of FISH results should be performed by suitably qualified staff, consistent with professional standards of practice, and should take into consideration other relevant test results, clinical and diagnostic information.
This device is intended for laboratory professional use only.
Failure to adhere to the protocol may affect the performance and lead to false positive/negative results.

Principles of the Test

Fluorescence *in situ* hybridisation (FISH) is a technique that allows DNA sequences to be detected on metaphase chromosomes or in interphase nuclei from fixed cytogenetic samples. The technique uses DNA probes that hybridise to entire chromosomes or single unique sequences, and serves as a powerful adjunct to G-banded cytogenetic analysis. This technique can now be applied as an essential investigative tool within prenatal, haematological and solid tumour chromosomal analysis. Target DNA, after fixation and denaturation, is available for annealing to a similarly denatured, fluorescently labelled DNA probe, which has a complementary sequence. Following hybridisation, unbound and non-specifically bound DNA probe is removed and the DNA is counterstained for visualisation. Fluorescence microscopy then allows the visualisation of the hybridised probe on the target material.

Probe Information

The *PML* (*promyelocytic leukaemia*) gene is located at 15q24.1 and the *RARA* (*retinoic acid receptor alpha*) gene is located at 17q21.2. The translocation t(15;17)(q24;q21) gives rise to the *PML::RARA* fusion gene and is the diagnostic hallmark of acute promyelocytic leukaemia (APL).

This FAST PML/RARα FISH probe allows rapid detection of the rearrangement, with only one hour of hybridisation required.

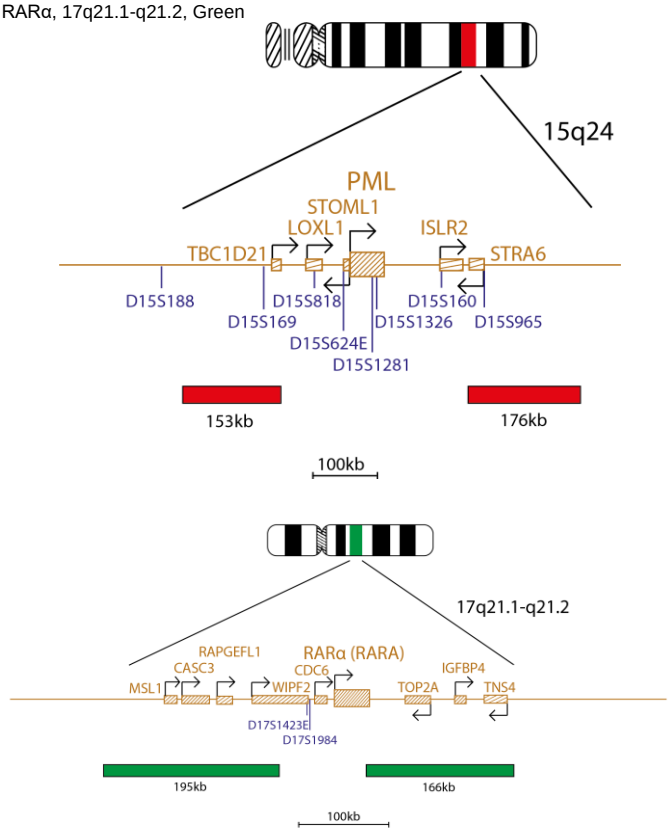
The *PML::RARA* fusion gene is created by the t(15;17)(q24;q21) translocation, found in more than 90% of cases of APL, a leukaemia that comprises 5-8% of cases of acute myeloid leukaemia (AML)[1,2]. In a subset of cases, variant *RARA* translocations can be observed. Known fusion partners include *NPM1* at 5q35, *NUMA1* at 11q13, *ZBTB16* (*PLZF*) at 11q23, *STAT5B* at 17q21, *PRKAR1A* at 17q24, *FIP1L1* at 4q12 and *BCOR* at Xp11[3,4,5].

PML and *RARA* have both been implicated in normal haematopoiesis. *PML* possesses growth suppressor and proapoptotic activity whereas *RARA* is a transcription factor that mediates the effect of retinoic acid at specific response elements[6]. PML::RARA fusion protein behaves as an altered retinoic acid receptor with an ability of transmitting oncogenic signaling[7].

Immediate treatment of APL patients is critical, due to fatal coagulation disorders and life-threatening haemorrhage in diagnosis. Prior to the introduction of all-trans-retinoic-acid (ATRA) and arsenic trioxide (ATO) in APL treatment protocols, the disease had a poor prognosis; however, since the introduction of these therapies, the overall survival rate has improved dramatically, with nearly 90%[5] of patients cured. Patients with variant *RARA* translocations show variable sensitivity to treatment, with some patients showing resistance to treatment protocols[3,5]. It is therefore important to differentiate between APL patients with *PML::RARA* fusion and those patients with variant *RARA* translocations.

Probe Specification

PML, 15q24 Red
RARα, 17q21.1-q21.2, Green



The PML probe mix, labelled in red, consists of a 153kb probe centromeric to the *PML* gene that covers the marker D15S169 and a 176kb probe telomeric to the *PML* gene that covers the marker D15S965. The *RARα* (*RARA*) probe mix, labelled in green, consists of a 195kb probe centromeric to the *RARα* (*RARA*) gene that spans the *CASC3* gene and a 166kb probe that covers the telomeric end of the *RARα* (*RARA*) gene as well as the *TOP2A*, *IGFBP4* and *TNS4* genes.

Materials Provided

Probe: 50µl per vial (5 tests), 100µl per vial (10 tests)
The probes are provided premixed in hybridisation solution (<65% formamide; <20mg dextran sulfate; <10% of 20x saline-sodium citrate (SSC)) and are ready to use.

Counterstain: 150µl per vial (15 tests)
The counterstain is DAPI Antifade ES (0.125µg/ml DAPI (4,6-diamidino-2-phenylindole) in glycerol-based mounting medium).

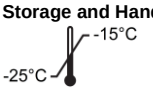
Warnings and Precautions

1. For *in vitro* diagnostic use. For laboratory professional use only.
2. Probe mixtures contain formamide, which is a teratogen; do not breathe fumes or allow skin contact. Handle with care; wear gloves and a lab coat.
3. Handle DAPI with care; wear gloves and a lab coat.
4. Do not use if the vial(s) are damaged, or the vial contents are compromised in any way.
5. Follow local disposal regulations for your location along with recommendations in the Safety Data Sheet to determine the safe disposal of this product. This also applies to damaged test kit contents.
6. Dispose of all used reagents and any other contaminated disposable materials following procedures for infectious or potentially infectious waste. It is the responsibility of each laboratory to handle solid and liquid waste according to their nature and degree of hazardousness and to treat and dispose of them (or have them treated and disposed of) in accordance with any applicable regulations.
7. Operators must be capable of distinguishing the colours red, blue, and green.
8. Failure to adhere to the outlined protocol and reagents may affect the performance and lead to false positive/negative results.
9. The probe should not be diluted or mixed with other probes.
10. Failure to use 10µl of probe during the pre-denaturation stage of the protocol may affect the performance and lead to false positive/negative results.
11. All products should be validated before use.
12. Internal controls should be carried out by using unaffected cell populations in testing samples.

Temperature Definitions

- -20°C / Frozen / In the Freezer: -25°C to -15°C
- 37°C: +37°C ± 1°C
- 72°C: +72°C ± 1°C
- 75°C: +75°C ± 1°C
- Room Temperature (RT): +15°C to +25°C

Storage and Handling



The kit should be stored between -25ºC to -15ºC in a freezer until the expiry date indicated on the kit label. The probe and counterstain vials must be stored in the dark.

The FISH probe, DAPI Antifade ES counterstain, and Hybridisation Solution remain stable throughout the freeze-thaw cycles experienced during normal use (where one cycle constitutes the vial's removal from and replacement into the freezer) - 5 cycles for the 50µl (5 tests) vial of FISH probe, 10 cycles for the 100µl (10 tests) vial of FISH probe, and 15 cycles for the 150µl (15 tests) vial of counterstain. Exposure to light should be minimised and avoided wherever possible. Store components in the light proof container provided. Components used and stored under conditions other than those stated on the labelling may not perform as expected and may adversely affect the assay results. All efforts must be made to limit exposure to light and temperature changes.

Equipment and Materials Necessary but not Supplied

Calibrated equipment must be used:

1. Hotplate (with a solid plate and accurate temperature control up to 80ºC)
2. Calibrated variable volume micropipettes and tips range 1µl - 200µl
3. Water bath with accurate temperature control at 37ºC and 72ºC
4. Microcentrifuge tubes (0.5ml)
5. Fluorescence microscope (Please see Fluorescence Microscope Recommendation section)
6. Phase contrast microscope
7. Clean plastic, ceramic or heat-resistant glass Coplin jars
8. Forceps
9. Calibrated pH meter (or pH indicator strips capable of measuring pH 6.5 – 8.0)
10. Humidified container
11. Fluorescence grade microscope lens immersion oil
12. Bench top centrifuge
13. Microscope slides
14. 24x24mm coverslips
15. Timer
16. 37ºC incubator
17. Rubber solution glue
18. Vortex mixer
19. Graduated cylinders
20. Magnetic stirrer
21. Calibrated thermometer

Optional Equipment not Supplied

1. Cytogenetic drying chamber

Reagents Needed but not Supplied

1. 20x saline-sodium citrate (SSC) Solution
2. 100% Ethanol
3. Tween-20
4. 1M Sodium hydroxide (NaOH)
5. 1M Hydrochloric acid (HCl)
6. Purified water

Fluorescence Microscope Recommendation

Use a 100-watt mercury lamp or equivalent and oil immersion plan apochromat objectives 60/63x or 100x for optimal visualisation. The fluorophores used in this probe set will excite and emit at the following wavelengths:

Fluorophore	$Excitation_{max}$ [nm]	$Emission_{max}$ [nm]
Green	495	521
Red	596	615

Ensure appropriate excitation and emission filters that cover the wavelengths listed above are fitted to the microscope. Use a triple bandpass DAPI/green spectrum/red spectrum filter or a dual bandpass green spectrum/red spectrum filter for optimal simultaneous visualisation of the green and red fluorophores.
Check the fluorescence microscope before use to ensure it is operating correctly. Use immersion oil that is suitable for fluorescence microscopy and formulated for low auto fluorescence. Avoid mixing DAPI antifade with microscope immersion oil as this will obscure signals. Follow manufacturers' recommendations in regards to the life of the lamp and the age of the filters.

Sample Preparation

The kit is designed for use on Carnoy's solution (3:1 methanol/acetic acid) fixed haematologically-derived cell suspensions, that are prepared according to the laboratory or institution guidelines. Prepare air dried samples on microscope slides according to standard cytogenetic procedures. The AGT *Cytogenetics Laboratory Manual* contains recommendations for specimen collection, culturing, harvesting and for slide making[8].

Solution Preparation

Ethanol Solutions

Dilute 100% ethanol with purified water using the following ratios and mix thoroughly:

- 70% Ethanol - 7 parts 100% ethanol to 3 parts purified water
- 85% Ethanol - 8.5 parts 100% ethanol to 1.5 parts purified water

Store the solutions for up to 6 months at room temperature in an airtight container.

2xSSC Solution

Dilute 1 part 20xSSC Solution with 9 parts purified water and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

0.4xSSC Solution

Dilute 1 part 20xSSC Solution with 49 parts purified water and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

2xSSC, 0.05% Tween-20 Solution

Dilute 1 part 20xSSC Solution with 9 parts purified water. Add 5µl of Tween-20 per 10ml and mix thoroughly. Check pH and adjust to pH 7.0 using NaOH or HCl as required. Store the solution for up to 4 weeks at room temperature in an airtight container.

***FAST* FISH Protocol – One (1) hour hybridisation**

(Note: Ensure that exposure of the probe and counterstain to laboratory lights is limited at all times).

Slide Preparation

1. Spot the cell sample onto a glass microscope slide. Allow to dry. (**Optional, if using a cytogenetic drying chamber**: The chamber should be operated at approximately 25°C and 50% humidity for optimal cell sample spotting. If a cytogenetic drying chamber is not available, use a fume hood as an alternative).
2. Immerse the slide in 2xSSC for 2 minutes at room temperature (RT) without agitation.
3. Dehydrate in an ethanol series (70%, 85% and 100%), each for 2 minutes at RT.
4. Allow to dry.

Pre-Denaturation

5. Remove the probe from the freezer and allow it to warm to RT. Briefly centrifuge tubes before use.
6. Ensure that the probe solution is uniformly mixed with a pipette.
7. Remove 10µl of probe per test, and transfer it to a microcentrifuge tube. Quickly return the remaining probe to the freezer.
8. Place the probe and the sample slide to prewarm on a 37ºC (+/- 1ºC) hotplate for 5 minutes.
9. Spot 10µl of probe mixture onto the cell sample and carefully apply a coverslip. Seal with rubber solution glue and allow the glue to dry completely.

Denaturation

10. Denature the sample and probe simultaneously by heating the slide on a hotplate at 75ºC (+/- 1ºC) for 2 minutes.

Hybridisation

11. Place the slide in a humid, lightproof container at 37ºC (+/- 1ºC) for one (1) hour.

Post-Hybridisation Washes

12. Remove the DAPI from the freezer and allow it to warm to RT.
13. Remove the coverslip and all traces of glue carefully.
14. Immerse the slide in 0.4xSSC (pH 7.0) at 72ºC (+/- 1ºC) for 2 minutes without agitation.
15. Drain the slide and immerse it in 2xSSC, 0.05% Tween-20 at RT (pH 7.0) for 30 seconds without agitation.
16. Drain the slide and apply 10µl of DAPI antifade onto each sample.
17. Cover with a coverslip, remove any bubbles and allow the colour to develop in the dark for 10 minutes.

18. View with a fluorescence microscope (see **Fluorescence Microscope Recommendation**).

Standard FISH Protocol – Overnight hybridisation
(Note: Ensure that exposure of the probe and counterstain to laboratory lights is limited at all times).

Slide Preparation
1. Spot the cell sample onto a glass microscope slide. Allow to dry. (**Optional, if using a cytogenetic drying chamber**: The chamber should be operated at approximately 25°C and 50% humidity for optimal cell sample spotting. If a cytogenetic drying chamber is not available, use a fume hood as an alternative).
2. Immerse the slide in 2xSSC for 2 minutes at room temperature (RT) without agitation.
3. Dehydrate in an ethanol series (70%, 85% and 100%), each for 2 minutes at RT.
4. Allow to dry.

Pre-Denaturation
5. Remove the probe from the freezer and allow it to warm to RT. Briefly centrifuge tubes before use.
6. Ensure that the probe solution is uniformly mixed with a pipette.
7. Remove 10µl of probe per test, and transfer it to a microcentrifuge tube. Quickly return the remaining probe to the freezer.
8. Place the probe and the sample slide to prewarm on a 37ºC (+/- 1ºC) hotplate for 5 minutes.
9. Spot 10µl of probe mixture onto the cell sample and carefully apply a coverslip. Seal with rubber solution glue and allow the glue to dry completely.

Denaturation
10. Denature the sample and probe simultaneously by heating the slide on a hotplate at 75ºC (+/- 1ºC) for 2 minutes.

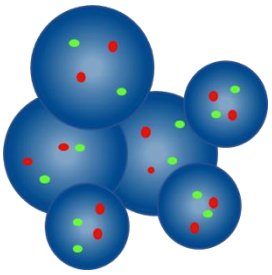
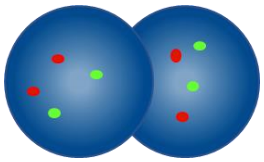
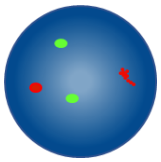
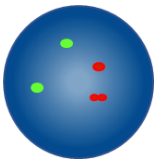
Hybridisation
11. Place the slide in a humid, lightproof container at 37ºC (+/- 1ºC) overnight.

Post-Hybridisation Washes
12. Remove the DAPI from the freezer and allow it to warm to RT.
13. Remove the coverslip and all traces of glue carefully.
14. Immerse the slide in 0.4xSSC (pH 7.0) at 72ºC (+/- 1ºC) for 2 minutes without agitation.
15. Drain the slide and immerse it in 2xSSC, 0.05% Tween-20 at RT (pH 7.0) for 30 seconds without agitation.
16. Drain the slide and apply 10µl of DAPI antifade onto each sample.
17. Cover with a coverslip, remove any bubbles and allow the colour to develop in the dark for 10 minutes.
18. View with a fluorescence microscope (see **Fluorescence Microscope Recommendation**).

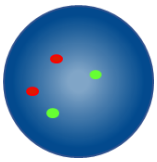
Procedural Recommendations
1. Baking or ageing of slides may reduce signal fluorescence.
2. Hybridisation conditions may be adversely affected by the use of reagents other than those provided or recommended by Cytocell Ltd.
3. Use a calibrated thermometer for measuring temperatures of solutions, waterbaths and incubators as these temperatures are critical for optimum product performance.
4. The wash concentrations, pH and temperatures are important as low stringency can result in non-specific binding of the probe and too high stringency can result in a lack of signal.
5. Incomplete denaturation can result in lack of signal and over denaturation can also result in non-specific binding.
6. Over hybridisation can result in additional or unexpected signals.
7. Users should optimise the protocol for their own samples prior to using the test for diagnostic purposes.
8. Suboptimal conditions may result in non-specific binding that may be misinterpreted as a probe signal.

Interpretation of Results
Assessing Slide Quality
The slide should not be analysed if:
- Signals are too weak to analyse in single filters - in order to proceed with analysis, signals should appear bright, distinct and easily evaluable
- There are high numbers of clumped/overlapping cells obstructing the analysis
- >50% of the cells are not hybridised
- There is excess of fluorescent particles between cells and/or a fluorescent haze that interferes with the signals - in optimal slides the background should appear dark or black and clean
- Cell nucleus borders cannot be distinguished and are not intact

Analysis Guidelines
- Two analysts should analyse and interpret each sample. Any discrepancies should be resolved by assessment by a third analyst
- Each analyst should be suitably qualified according to recognised national standards
- Each analyst should score independently 100 nuclei for each sample. The first analyst should start the analysis from the left side of the slide and the second analyst from the right one
- Each analyst should document their results in separate sheets
- Analyse only intact nuclei, not overlapped or crowded nuclei or nuclei covered by cytoplasmic debris or high degree of autofluorescence
- Avoid areas where there is excess of cytoplasmic debris or non-specific hybridisation
- Signal intensity may vary, even with a single nucleus. In such cases, use single filters and/or adjust the focal plane
- In suboptimal conditions signals may appear diffuse. If two signals of the same colour touch each other, or the distance between them is no greater than two signal widths, or when there is a faint strand connecting the two signals, count as one signal
- When analysing dual-colour breakapart probes, if there is a gap between the red and green signal no greater than 2 signals width apart, count as not rearranged/fused signal
- When analysing three-colour breakapart probes, if there is a gap between any of the 3 signals (red, green, blue) no greater than 2 signals width apart, count as not rearranged/fused signal
- If in doubt about whether a cell is analysable or not, then do not analyse it

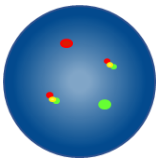
Analysis Guidelines	
	Do not count – nuclei are too close together to determine boundaries
	Do not count overlapping nuclei – all areas of both nuclei are not visible
	Count as two red signals and two green signals – one of the two red signals is diffuse
	Count as two red signals and two green signals – the gap in one red signal is less than two signal widths

Expected Results
Expected Normal Signal Pattern



In a normal cell, two red and two green signals (2R2G) are expected.

Expected Abnormal Signal Patterns



In an a cell with t(15;17)(q24.1;q21) translocation, one red, one green and two fusions (1R1G2F) are expected.

Other signal patterns are possible in aneuploid/unbalanced specimens.

Known Relevant Interferences / Interfering Substances
No known relevant interferences / interfering substances.

Known Cross-Reactivity
No known cross-reactivity.

Serious Incident Reporting
For a patient/user/third party in the European Union and in countries with identical regulatory regime (Regulation (EU) 2017/746 on *In vitro* Diagnostic Medical Devices); if, during the use of this device or as a result of its use, a serious incident has occurred, please report it to the Manufacturer and to your National Competent Authority.
For serious incidents in other countries, please report it to the Manufacturer and, if applicable, to your National Competent Authority.
Manufacturer vigilance contact: vigilance@ogt.com
For EU National Competent Authorities, a list of vigilance contact points can be found at:
https://health.ec.europa.eu/medical-devices-sector/new-regulations/contacts_en

Specific Performance Characteristics
Analytical Specificity
Analytical specificity is defined as the percentage of signals that hybridise to the correct locus and no other location. Four chromosomal loci in each of twenty metaphase cells from each of the five samples were analysed, giving 400 data points. The location of each hybridised probe was mapped and the number of metaphase chromosome FISH signals that hybridised to the correct locus was recorded.

The analytical specificity of each probe in the kit was calculated as the number of metaphase chromosome FISH signals hybridised to the correct locus divided by the total number of metaphase chromosome FISH signals hybridised, this result was multiplied by 100, expressed as a percentage and given with a 95% confidence interval.

Table 1. Analytical Specificity for the *FAST* PML/RARα (RARA) Translocation, Dual Fusion Probe

Target	Number of metaphase chromosomes hybridised	Number of correctly hybridised loci	Analytical Specificity	95% Confidence Interval
15q24.1	200	200	100%	98.12% -100%
17q21.1-17q21.2	200	200	100%	98.12% -100%

Analytical Sensitivity
Analytical sensitivity is the percentage of scoreable interphase cells with the expected normal signal pattern. A minimum of 100 interphase cells were analysed for each of 25 fixed cell suspensions from bone marrow and 25 fixed cell suspensions from peripheral blood using the fast hybridisation method, and 25 fixed cell suspensions from bone marrow, using an overnight hybridisation method. This resulted in a minimum of 2500 nuclei scored for peripheral blood samples and 5000 nuclei scored for bone marrow samples. The sensitivity data was analysed based on the percentage of cells showing a normal expected signal pattern and expressed as a percentage with a 95% confidence interval.

Table 2. Analytical Sensitivity for the *FAST* PML/RARα (RARA) Translocation, Dual Fusion Probe

Sample Type	Sensitivity Criteria	Sensitivity Result
Bone-Marrow-Fast hybridization	>95%	98.80% (97.96 – 99.63%)
Bone-Marrow-Overnight hybridization	>95%	98.52% (97.76-99.28%)
Peripheral Blood- Fast hybridization	>95%	99.31% (98.66 – 100.00%)

Characterisation of Normal Cut-off Values
The normal cut-off is defined as the percentage of cells exhibiting a false positive signal pattern at which an individual would be considered normal and not consistent with a clinical diagnosis A minimum of 100 interphase cells were analysed for each of 25 fixed cell suspensions from bone marrow and 25 fixed cell suspensions from peripheral blood using the fast hybridisation method, and 25 fixed cell suspensions from bone marrow, using an overnight hybridisation method. This resulted in a minimum of 2500 nuclei scored for peripheral blood samples and 5000 nuclei scored for bone marrow samples.

The cut-off value was determined using the β-inverse (BETAINV) function in MS Excel. It was calculated as the percentage of interphase cells showing a false positive signal pattern using the upper bound of a one-sided 95% confidence interval of the binomial distribution in a normal patient sample.

Table 3. Characterisation of Normal Cut-off Values for the *FAST* PML/RARα (RARA) Translocation, Dual Fusion Probe

Sample Type	Cut-off Result
Bone Marrow - Fast hybridisation	2.71%
Bone Marrow - Overnight hybridisation	3.44%
Peripheral Blood - Fast hybridisation	4.36%

Laboratories must verify cut-off values using their own data[9,10].

Precision
The precision of this product has been measured in terms of intra-day precision (sample-to-sample), inter-day precision (day-to-day) and single-site inter-lot precision (lot-to-lot).

Two samples per hybridisation method were used to assess the precision of this product: a negative bone marrow and a low positive marrow. The low positive bone marrow sample (2-4x the product's cut-off), was created by spiking the normal bone marrow sample with a known positive bone marrow sample, and was used to challenge the product around the established cut-off.

To establish the inter-day and intra-day precision, the samples were evaluated over ten non-consecutive dates and to establish the lot-to-lot precision, three lots of the product were evaluated on three replicates of the same samples. The results were presented as the overall agreement with the predicted negative class (for the negative samples).

Table 4. Reproducibility and Precision for the *FAST* PML/RARα (RARA) Translocation, Dual Fusion Probe

Variable	Sample type	Agreement
Intra-day (sample to sample) & inter-day (day to day) reproducibility	Bone Marrow Negative	100%
	Bone Marrow Low Positive	100%
Lot-to-lot reproducibility	Bone Marrow Negative	100%
	Bone Marrow Low Positive	100%

Clinical Performance
To ensure that the product detects intended rearrangements, clinical performance was established over one study on representative samples of the intended population for the product: residual methanol/acetic acid fixed haematologically derived material. The sample size was 136 specimens, with a population of 43 positive specimens and 93 negative specimens. The results were compared to the known status of the sample, as identified by a comparator method. The concordance/discordance of results was found to meet the acceptance criteria for this study.

The results of these tests were analysed in order to provide clinical sensitivity, clinical specificity and false positive rate (FPR) values for positive signals, using a one-dimensional approach.

Table 5. Clinical Performance for the *FAST* PML/RARα (RARA) Translocation, Dual Fusion Probe

Variable	Result
Clinical Sensitivity (true positive rate, TPR)	98.93%
Clinical Specificity (true negative rate, TNR)	99.58%
False Positive rate (FPR) = 1 – Specificity	0.42%

Summary of Safety and Performance (SSP)
The SSP shall be made available to the public via the European database on medical devices (Eudamed), where it is linked to the Basic UDI-DI.
Eudamed URL: https://ec.europa.eu/tools/eudamed
Basic UDI-DI: 50558449LPH064JR

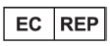
If Eudamed is not fully functional, the SSP shall be made available to the public upon request by emailing SSP@ogt.com.

Additional Information
For additional product information, please contact the CytoCell Technical Support Department.
T: +44 (0)1223 294048
E: techsupport@cytocell.com
W: www.ogt.com

References

1. Swerdlow, *et al* (eds). WHO Classification of Tumours of Haematopoietic and Lymphoid Tissue, Lyon, France, 4th edition, IARC, 2017
2. Campbell, *et al*. Biomed Research International. 2013;2013:1-5.
3. Creutzig, *et al*. Blood. 2012;120(16):3187-3205.
4. Zhang, *et al*. Blood Reviews. 2015;29(2):101-125.
5. Tomita, *et al*. International Journal of Haematology. 2013;97(6):717-725.
6. Grimwade, *et al*. Blood. 2000;96(4):1297-1308.
7. Lo-Coco, Hasa. Best practice & research. Clinical haematology. 2014;27(1):3-9.
8. Arsham, MS., Barch, MJ. and Lawce HJ. (eds). (2017) The AGT Cytogenetics Laboratory Manual. New Jersey: John Wiley & Sons Inc.
9. Mascarello JT, Hirsch B, Kearney HM, *et al*. Section E9 of the American College of Medical Genetics technical standards and guidelines: fluorescence in situ hybridization. Genet Med. 2011;13(7):667-675.
10. Wiktor AE, *et al*. Preclinical validation of fluorescence in situ hybridization assays for clinical practice. Genetics in Medicine. 2006;8(1):16–23.

Symbols Glossary

EN ISO 15223-1:2021 - "Medical devices - Symbols to be used with information to be supplied by the manufacturer - Part 1: General requirements" (© International Organization for Standardization)		
Symbol	**Title**	**Reference Number(s)**
	**en:** Manufacturer	5.1.1
	**en:** Authorized representative in the European Community/European Union	5.1.2
	**en:** Use-by date	5.1.4
	**en**: Batch code	5.1.5
	**en:** Catalogue number	5.1.6
	**en:** Keep away from sunlight	5.3.2
	**en:** Temperature limit	5.3.7
	**en:** Consult instructions for use	5.4.3
 ogt.com/IFU	**en:** Consult electronic instructions for use	5.4.3
	**en:** Caution	5.4.4
	**en:** *In vitro* diagnostic medical device	5.5.1
	**en:** Contains sufficient for <n> tests	5.5.5
	**en:** Unique Device Identifier	5.7.10
EDMA symbols for IVD reagents and components, October 2009 revision		
Symbol	**Title**	**Reference Number(s)**
	**en:** Contents (or contains)	N/A

Patents and Trademarks
CytoCell is a registered trademark of Cytocell Limited.

Cytocell Limited
Oxford Gene Technology
418 Cambridge Science Park
Milton Road
CAMBRIDGE
CB4 0PZ
UNITED KINGDOM

T: +44 (0)1223 294048
F: +44 (0)1223 294986
E: probes@cytocell.com
W: www.ogt.com

Sysmex Europe SE
Bornbarch 1
22848 Norderstedt
GERMANY

T: +49 40 527260
W: www.sysmex-europe.com

IFU Version History
V001.00 2023-01-25: New IFU for Regulation (EU) 2017/746